RESEARCH



Differential expression of serum mir-363-3p in patients with polycystic ovary syndrome and its predictive value for their pregnancy



Min Sang¹, Ying Yu¹, Zhi Zhou¹, Yaqi Zhang² and Haiping Chang^{3*}

Abstract

Background This study aimed to investigate the expression of serum miR-363-3p in patients with polycystic ovary syndrome (PCOS) and its predictive value for pregnancy after ovulation induction therapy.

Methods The expression of serum miR-363-3p was detected by Reverse transcription quantitative polymerase chain reaction (RT-qPCR). PCOS patients were treated with ovulation induction therapy, and after the successful pregnancy was confirmed, they were followed up for 1 year in outpatient department to record the pregnancy outcomes of the patients. The Pearson correlation coefficient was used to evaluate the correlation between the expression level of miR-363-3p and biochemical indicators of PCOS patients. Logistic regression analysis was used to analyze the risk factors of pregnancy failure after ovulation induction therapy.

Results The serum level of miR-363-3p in PCOS group was significantly lower than that in control group. Compared with the control group, both pregnant and non-pregnant groups had lower miR-363-3p levels, while the non-pregnant group had a greater reduction in miR-363-3p levels than the pregnant group. Low levels of miR-363-3p showed high accuracy in distinguishing pregnant and non-pregnant patients. Logistic regression analysis showed that high levels of luteinizing hormone, testosterone (T), prolactin (PRL) and low level of miR-363-3p were independent risk factors for pregnancy failure after ovulation induction in PCOS patients. Additionally, compared with pregnancy outcomes of healthy women, the incidence of premature delivery, macrosomia, and gestational diabetes in PCOS patients increased.

Conclusions The expression of miR-363-3p in PCOS patients was reduced and correlated with abnormal hormone levels, suggesting that miR-363-3p may be involved in the occurrence and development of PCOS.

Keywords Polycystic ovarian syndrome, Sex hormone, MiR-363-3p, Pregnancy outcome

Haipingchang1975@126.com

- University (Dabie Mountain Regional Medical Center),
- Huanggang 438000, Hubei, P.R. China

²Scientific Research and Teaching Department, Huanggang Central

Hospital of Yangtze University (Dabie Mountain Regional Medical Center), Huanggang 438000, Hubei, P.R. China

³Department of Gynecology, Huanggang Central Hospital of Yangtze University (Dabie Mountain Regional Medical Center), No.126 Qi'an Avenue, Huangzhou District, Huanggang 438000, Hubei, P.R. China



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence:

Haiping Chang

¹Reproductive Medicine Center, Huanggang Central Hospital of Yangtze

Background

Polycystic ovary syndrome (PCOS) is a common endocrine system disease in women of childbearing age, and it is also a research hotspot and focal issue in the entire field of clinical reproductive medicine [1]. There are some individual differences in PCOS patients, but the clinical manifestations are mainly concentrated in three aspects of sparse ovulation, polycystic ovarian changes and hyperandrogenism [2]. According to research statistics, the incidence of PCOS in women of childbearing age is about 10%, and infertility caused by PCOS accounts for about 60% of all infertile patients [3]. With the in-depth study of PCOS, people have also discovered the longterm impact of PCOS on women. In fact, PCOS can not only affect the women's reproductive function, but also lead to metabolic disorders, such as hyperinsulinemia and insulin resistance, thus increasing the risk of secondary diabetes, cardiovascular disease, endometrial cancer, which has a negative impact on women's health [4, 5].

MicroRNAs are a kind of non-coding RNAs with a length of 18-24 nucleotides, which is widely distributed in eukaryotes and usually play a negative regulatory role in vivo [6]. In recent years, there are more and more studies on miRNA in PCOS, and the research samples include serum, follicular fluid, ovarian granulosa cells and PCOS animal models [7]. MiRNA family is large and numerous. At present, only part of miRNA has been explored in PCOS. After analyzing the data of granular cell microarray in PCOS patients, Wang et al. found that miR-486-5p was involved in regulating the growth and development of follicles [8]. Another study found that there were differentially expressed miRNAs in follicular fluid, such as miR-135 and miR-32, which are potential new biomarkers of PCOS [9]. MiR-363-3p, located on the human X chromosome, is one of the members of the miR-92a family. At present, the function of miR-363-3p mainly focused on tumor immunity [10]. Studies have shown that miR-363-3p can promote the occurrence of gastric cancer by down-regulating the expression of tumor suppressor gene MBP-1 [11]. In addition, the level of miR-363-3p in vivo is related to the invasion and metastasis of head and neck squamous cell carcinoma by acting on tumor invasion and metastasis-related proteins [12]. A recent study found that the decrease of miR-363-3p in obese people with metabolic syndrome was associated with the increase of serum cholesterol [13]. Liu et al. found that there were abnormal expressions of five miRNAs in ovarian endometriosis, including miR-363-3p, and their expression in ectopic endometrium group was significantly reduced compared with normal endometrium tissue [14]. Another study showed that the expression of miR-363-3p in plasma exosomes was inconsistent between PCOS patients and healthy people, showing a downregulation trend in the PCOS group [15].

Nevertheless, the current research on the relationship between miR-363-3p and PCOS is still unclear. The purpose of this study was to analyze the expression level of miR-363-3p in PCOS patients, the relationship between miR-363-3p and the patient's blood biochemical indicators, and the predictive value of miR-363-3p for pregnancy in PCOS patients.

Methods

Inclusion of the study population

A total of 126 infertile patients due to PCOS admitted to Huanggang Central Hospital of Yangtze University were selected. The diagnosis of PCOS was in accordance with the 2003 Rotterdam criteria [16]. Inclusion criteria: (1) Rare ovulation and/or anovulation. The standard of rare ovulation is menstrual cycle>35 days. (2) The clinical manifestations and/or biochemical signs of hyperandrogenism, among which hyperandrogenism is based on the elevated total testosterone level measured in laboratory. The clinical features of hyperandrogenism may be acne and hirsutism (excluding other disorders that can lead to hyperandrogenism, such as congenital adrenal hyperplasia, androgen-secreting tumors, and Cushing's syndrome). (3) Polycystic ovarian changes: ≥ 12 follicles with a diameter of 2-9 mm in one or both ovaries, and/or ≥10mL in ovarian volume. PCOS can be diagnosed when patients meet any two of the above three criteria. The control group consisted of 110 infertile women caused by male factors, with normal menstruation, no clinical manifestations of androgen excess, normal ovulation and normal morphology of bilateral ovary by ultrasound examination. Exclusion criteria: (1) History of ovarian surgery or radiotherapy and chemotherapy, endometriosis, uterine fibroids, endometrial polyps, uterine malformations, intrauterine adhesions, various genetic diseases, and acute inflammation. (2) No history of hormone medication in the past three months. All subjects were unrelated Han Chinese from the same geographical area. This study was performed in accordance with the principles of the Declaration of Helsinki. The experimental scheme complies with the requirements of the Ethics Committee of Huanggang Central Hospital of Yangtze University. The study is approved by the Ethics Committee of Huanggang Central Hospital of Yangtze University, and the patients and their families signed the informed consent.

During the subject's natural menstrual cycle or during the period of progesterone-induced withdrawal bleeding in amenorrhea (follicular phase: days 3–5 of menstruation), after a 10-hour overnight fasting period, a 5 mL venous blood sample was drawn the next morning. The serum levels of luteinizing hormone, follicle-stimulating hormone (FSH), prolactin (PRL), estradiol (E2), progesterone (P), total testosterone (TT), fasting blood glucose (FBG) and fasting insulin were measured by chemiluminescence immunoassay. Total cholesterol (TC), triglyceride (TG), high-density lipoprotein cholesterol (HDL-C) and low-density lipoprotein cholesterol (LDL-C) were determined by colorimetry. The homeostasis model of insulin resistance (HOMA-IR) index was used to evaluate the insulin resistance of patients. HOMA-IR=FBG level (mmol/L) \cdot fasting insulin level (mIU/L) / 22.5.

Treatment methods and sample collection

All patients received ovulation induction therapy. Clomiphene citrate, 50 mg/d, was taken orally from the third day of menstrual cycle or withdrawal bleeding of progesterone for 5 consecutive days. On the 8th day of menstruation, human menopausal urinary gonadotropin (75 IU/d) was injected intramuscularly, and the duration of administration was determined according to follicular development. When the diameter of the follicle is \geq 18 mm and the urine LH (+), the intramuscular injection of chorionic gonadotropin is started, and the intercourse was performed during the ovulation. Fasting peripheral venous blood was taken 6–8 days after ovulation, and the supernatant was collected after centrifugation and stored in -80°C refrigerator for later use.

Table 1 Comparison of general data between PCOS group and
control group

Indicators	Controls	PCOS (n = 126)	Ρ
	(n = 110)		value
Age (Year)	28.75 ± 2.41	28.64 ± 2.56	0.426
BMI (kg/m ²)	23.41 ± 2.51	23.96 ± 2.72	0.103
SBP (mm Hg)	118.36 ± 13.14	119.73±12.98	0.108
DBP (mm Hg)	72.48±6.91	74.22 ± 6.82	0.465
FBG (mmol/L)	4.68 ± 0.46	5.01 ± 0.82	0.000
FINS (mIU/L)	7.16 ± 1.54	23.96 ± 4.91	0.000
HOMA-IR	1.57 ± 0.32	2.44 ± 0.95	0.000
LH (IU/L)	5.13 ± 1.75	12.31 ± 5.63	0.000
FSH (IU/L)	6.12 ± 1.81	7.88 ± 1.42	0.000
E ₂ (pmol/L)	173.76 ± 40.51	102.61±45.66	0.000
T (µg/L)	0.43 ± 0.20	0.89 ± 0.23	0.000
PRL (µg/L)	10.57 ± 4.27	13.85 ± 5.24	0.000
P (nmol/L)	0.81 ± 0.41	1.15 ± 0.49	0.000
AMH (ng/mL)	3.42 ± 1.36	9.59 ± 2.11	0.000
TC (mmol/L)	4.60 ± 0.36	4.95 ± 0.33	0.045
TG (mmol/L)	0.95 ± 0.13	0.92 ± 0.21	0.387
HDL-C (mmol/L)	1.60 ± 0.10	1.63 ± 0.08	0.174
LDL-C (mmol/L)	2.56 ± 0.14	2.52 ± 0.18	0.225

Abbreviations: PCOS: polycystic ovary syndrome; BMI: body mass index; SBP: systolic blood pressure; DBP: diastolic blood pressure; FBG: fasting blood glucose; FINS: fasting insulin; HOMA-IR: homeostasis model assessment of insulin resistance; LH: luteinizing hormone; FSH: follicle-stimulating hormone; E_2 : estradiol; T: testosterone; PRL: prolactin; P: progesterone; AMH: anti-mullerian hormone; TC: total cholesterol; TG: triglyceride; HDL-C: high-density lipoprotein cholesterol; LDL-C: low-density lipoprotein cholesterol. Data are presented as mean \pm standard deviation (SD).

Detection of serum miR-363-3p

Reverse transcription quantitative polymerase chain reaction (RT-qPCR) was used to detect the serum level of miR-363-3p. Total RNA in serum was extracted according to the instructions of TRIzol, and the purity of RNA was detected by spectrophotometer. RNA was reverse transcribed into cDNA by reverse transcription kit, and U6 was used as internal reference for amplification. All primers needed were synthesized and provided by GenePharma Company (Nanjing, China). Reaction conditions: 94°C, 3min, 1 cycle; 92°C, 30s, 60°C, 30s, 75°C, 30s, 40 cycles. Primers: miR-363-3p: 5'-CGCAAGAACATCTC-CAAG-3' (forward), 5'-CTCAACTGGTGTCGTGGA-3' (reverse); U6: 5'-CTCGCTTCGGCAGCACA-3' (forward), 5'-AACGCTTCACGAATTTGCGT-3' (reverse). After the reaction, the relative expression of serum miR-363-3p was calculated by $2^{-\Delta\Delta Ct}$ method.

Pregnancy judgment and follow-up analysis

Serum human chorionic gonadotropin (HCG) was measured on the 14th day after ovulation, and when the HCG level was significantly increased, it was defined as biochemical pregnancy. Clinical pregnancy was confirmed when the presence of a gestational sac was indicated by transvaginal ultrasound on day 30–35 after ovulation. The patients were followed up for one year through the outpatient clinic, and the pregnancy status of the patients was recorded during the follow-up period.

Statistical analysis

SPSS 20.0 was used for data processing. Counting data are represented by n for Chi-square test; measurement data conforming to normal distribution are represented by (mean \pm SD) for *t* test. Pearson test was used for correlation analysis. Logistic regression analysis was used to evaluate the risk factors of pregnancy failure after ovulation induction therapy in PCOS patients.

Results

General clinical conditions

The general clinical data and biochemical indicators of the control group and PCOS group are summarized in Table 1. It can be seen from this result that there were no significant differences in age, body mass index (BMI), systolic blood pressure (SDP), diastolic blood pressure (DBP), triglyceride (TG), high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C) between control group and PCOS group (P>0.05). Notably, in PCOS group, the levels of fasting blood glucose (FBG), fasting insulin (FINS), homeostasis model of insulin resistance (HOMA-IR), luteinizing hormone, follicle-stimulating hormone (FSH), estradiol (E2), testosterone (T), prolactin (PRL), progesterone

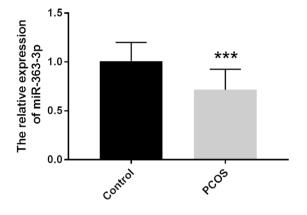


Fig. 1 The expression level of serum miR-363-3p in control and PCOS groups. Serum level of miR-363-3p in PCOS group was reduced in comparison to the control group. ***P<0.001

 Table 2
 Correlation between serum miR-363-3p level and

Indexes	Correlation with serum miR-363-3p level (r)	P value	
Age	-0.154	0.515	
BMI	-0.311	0.063	
SBP	-0.116	0.625	
DBP	-0.108	0.713	
FBG	-0.604	< 0.001	
FINS	-0.593	< 0.001	
HOMA-IR	-0.581	< 0.001	
LH	-0.635	< 0.001	
FSH	-0.670	< 0.001	
E ₂	-0.722	< 0.001	
Т	-0.757	< 0.001	
PRL	-0.594	< 0.001	
Р	-0.732	< 0.001	
AMH	-0.559	< 0.001	
TC	-0.401	0.031	
TG	-0.019	0.862	
HDL-C	-0.131	0.606	
LDL-C	-0.144	0.533	

Abbreviations: PCOS: polycystic ovary syndrome; BMI: body mass index; SBP: systolic blood pressure; DBP: diastolic blood pressure; FBG: fasting blood glucose; FINS: fasting insulin; LH: luteinizing hormone; FSH: follicle-stimulating hormone; E2: estradiol; T: testosterone; PRL: prolactin; P: progesterone; AMH: anti-mullerian hormone; HOMA-IR: homeostasis model assessment of insulin resistance; TC: total cholesterol; TG: triglyceride; HDL-C: high-density lipoprotein cholesterol

(P), anti-mullerian hormone (AMH), and total cholesterol (TC) were significantly higher than those in control group (P<0.05).

Table 3	Comparison of indicators between pregnant group and	
non-preg	nant group in PCOS patients	

Indicators	Pregnancy group (n=70)	Non-pregnan- cy group (n=56)	P value	
Age (Year)	28.39 ± 2.41	28.31 ± 2.49	0.656	
BMI (kg/m ²)	23.51 ± 2.86	23.52 ± 2.56	0.517	
SBP (mm Hg)	120.66±11.79	121.64 ± 12.12	0.791	
DBP (mm Hg)	73.05 ± 7.08	74.82 ± 6.93	0.153	
FBG (mmol/L)	4.99 ± 0.87	5.11 ± 0.81	0.051	
FINS (mIU/L)	9.22±1.78	12.96 ± 2.15	0.000	
HOMA-IR	2.02 ± 0.21	2.61 ± 0.37	0.000	
LH (IU/L)	10.27±2.26	16.02 ± 4.16	0.000	
FSH (IU/L)	7.01 ± 2.04	10.22 ± 3.13	0.000	
E ₂ (pmol/L)	171.16 ± 106.35	170.81 ± 104.34	0.756	
T (µg/L)	0.63 ± 0.15	1.02 ± 0.30	0.000	
PRL (µg/L)	13.80 ± 4.75	13.93 ± 5.59	0.891	
P (nmol/L)	1.23 ± 0.51	1.06 ± 0.52	0.015	
AMH (ng/mL)	6.86 ± 1.59	11.23 ± 2.01	0.000	
TC (mmol/L)	4.85 ± 0.34	4.90 ± 0.35	0.261	
TG (mmol/L)	0.91 ± 0.20	0.89 ± 0.23	0.279	
HDL-C (mmol/L)	1.64 ± 0.11	1.62 ± 0.07	0.356	
LDL-C (mmol/L)	2.59±0.21	2.61 ± 0.26	0.217	

Abbreviations: PCOS: polycystic ovary syndrome; BMI: body mass index; SBP: systolic blood pressure; DBP: diastolic blood pressure; FBG: fasting blood glucose; FINS: fasting insulin; LH: luteinizing hormone; FSH: follicle-stimulating hormone; E_2 : estradiol; T: testosterone; PRL: prolactin; P: progesterone; AMH: anti-mullerian hormone; HOMA-IR: homeostasis model assessment of insulin resistance; TC: total cholesterol; TG: triglyceride; HDL-C: high-density lipoprotein cholesterol; LDL-C: low-density lipoprotein cholesterol. Data are presented as mean±standard deviation (SD).

Serum mir-363-3p expression level and its correlation with clinical data of PCOS patients

The expression level of serum miR-363-3p in control group and PCOS group was detected by RT-qPCR. Compared with the control group, the relative expression level of miR-363-3p was significantly decreased (Fig. 1, P < 0.001). Additionally, Pearson correlation coefficient revealed that the expression of serum miR-363-3p was negatively correlated with FBG, FINS, HOMA-IR, LH, FSH, E2, T, PRL, P, AMH, TC (Table 2, P < 0.05).

Pregnancy status in PCOS patients after ovulation induction therapy

After systematic ovulation induction treatment, 70 of 126 PCOS patients were successfully conceived, and 56 failed. Patients who successfully conceived were classified as pregnant group, while patients who were not pregnant were classified as non-pregnant group. The clinical information of these two groups is summarized in Table 3. After analysis, it could be seen that the level of BMI, FINS, HOMA-IR, LH, FSH, T, P, and AMH of non-pregnant group is significantly higher than those in pregnant group (P<0.05).

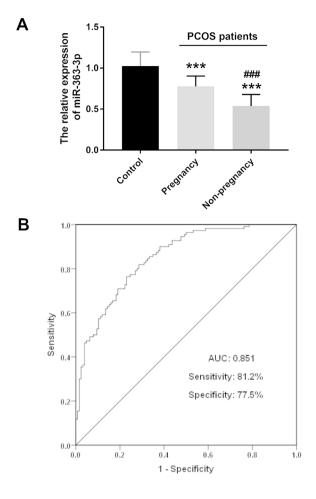


Fig. 2 Expression of miR-363-3p in PCOS pregnancy group and non-pregnancy group and its discriminative effect on pregnancy and non-pregnancy. **(A)** Compared with the control group, the miR-363-3p level of both PCOS pregnant group and PCOS non-pregnant group was significantly decreased, while the miR-363-3p level of PCOS non-pregnant group was lower than that of the pregnant group. **(B)** ROC analysis. MiR-363-3p has a good discrimination value between pregnant and non-pregnant people in PCOS patients. ****P* < 0.001 VS Control group; ###*P* < 0.001 VS PCOS pregnant group

Expression and diagnostic value of mir-363-3p in pregnancy and non-pregnancy groups

As shown in Fig. 2A, compared with the control group, both the pregnant group and the non-pregnant group had significantly lower miR-363-3p levels (P<0.001). In addition, the miR-363-3p levels in the non-pregnancy group was lower than that in the pregnant group (P<0.001). Furthermore, ROC analysis showed that miR-363-3p had a high diagnostic accuracy for distinguishing pregnant women from non-pregnant people in PCOS patients, with the area under the curve (AUC) of 0.851, the sensitivity and specificity of 81.2% and 77.5%, respectively (see Fig. 2B).

Table 4	Factors	affecting	pregnancy	failure ir	n PCOS p	atients
after indu	uction o	f excretior	۱			

Variables	OR	95% CI	Р	
			value	
Age	1.172	0.513-2.677	0.706	
BMI	0.821	0.340-1.982	0.660	
SBP	2.128	0.855-5.294	0.105	
DBP	1.520	0.637-3.629	0.346	
FBG	0.708	0.298-1.685	0.435	
FINS	1.754	0.738-4.166	0.203	
HOMA-IR	0.652	0.269-1.590	0.343	
LH	2.839	1.175–6.863	0.020	
FSH	2.502	0.974–6.428	0.059	
E2	0.946	0.404-2.214	0.899	
Т	3.689	1.872-9.253	0.009	
PRL	2.562	1.062-6.178	0.036	
Р	1.561	0.684-3.527	0.329	
AMH	1.228	0.542-2.782	0.622	
TC	0.720	0.296-1.749	0.468	
TG	1.345	0.601-3.009	0.490	
HDL-C	1.192	0.526-2.705	0.674	
LDL-C	1.165	0.494-2.747	0.727	
miR-363-3p	0.243	0.095-0.617	0.003	

Abbreviations: PCOS: polycystic ovary syndrome; BMI: body mass index; SBP: systolic blood pressure; DBP: diastolic blood pressure; FBG: fasting blood glucose; FINS: fasting insulin; LH: luteinizing hormone; FSH: follicle-stimulating hormone; E₂: estradiol; T: testosterone; PRL: prolactin; P: progesterone; AMH: anti-mullerian hormone; HOMA-IR: homeostasis model assessment of insulin resistance; TC: total cholesterol; TG: triglyceride; HDL-C: high-density lipoprotein cholesterol

Table 5 Comparison of pregnancy outcomes between control group and pregnancy group

Control group (n=110)	Pregnancy group (n=70)
3 (2.73%)	10 (14.29%) *
2 (1.82%)	4 (5.71%)
2 (1.82%)	9 (12.86%) *
4 (3.64%)	12 (17.14%) *
2 (1.82%)	1 (1.43%)
6 (5.45%)	6 (8.57%)
	(n = 110) 3 (2.73%) 2 (1.82%) 2 (1.82%) 4 (3.64%) 2 (1.82%)

Note: Data are expressed as n (%). *P<0.05

Possible factors influencing pregnancy failure after ovulation induction therapy in PCOS patients

Logistic regression analysis was used to evaluate the influencing factors of pregnancy success in PCOS patients. Table 4 showed that high levels of LH (OR: 2.839, 95% CI: 1.175–6.863, P=0.020), T (OR: 3.689, 95% CI: 1.872–9.253, P=0.009), PRL (OR: 2.562, 95% CI: 1.062–6.178, P=0.036) and low level of miR-363-3p (OR: 0.243, 95% CI: 0.095–0.617, P=0.003) were independent risk factors for unsuccessful pregnancy in PCOS patients after ovulation induction therapy.

Comparison of pregnancy outcomes between control and PCOS pregnancy groups

After the follow-up, the pregnancy outcomes and the incidence of related complications were summarized and analyzed. The results suggested that compared with the control group, the incidence of premature delivery, macrosomia and gestational diabetes in the PCOS pregnancy group was significantly enhanced (Table 5, P<0.05).

Discussion

PCOS is the most common endocrine disorder syndrome in women of childbearing age, which can not only lead to infertility, but also be associated with insulin resistance, obesity, diabetes and cardiovascular disease [17, 18]. Therefore, the monitoring and evaluating the treatment of PCOS is an important link to reduce the related long-term complications. Despite the long-term and detailed study, it is still difficult to clarify the etiology and pathogenesis of PCOS. At present, the combined effect of genetic and environmental factors is a recognized conclusion of the pathogenesis of PCOS [19]. In recent years, researchers on the etiology of PCOS have begun to pay attention to non-coding RNAs, especially miRNA. MiRNA participates in various biological activities of the body, and its expression is different in pathological conditions such as malignant tumor or endocrine and metabolic diseases. Similarly, miRNAs are involved in the metabolic processes of body substances, including the metabolism and transport of cholesterol and fatty acids, the regulation of pancreas islet function and the metabolism and transport of glucose [20, 21]. The above evidence indicates that miRNAs play an extraordinary role in human physiology and disease.

In this study, the expression level of serum miR-363-3p in PCOS group was significantly lower than that in the control group. The expression levels of serum miR-363-3p in this study were consistent with the results in plasma exosomes of PCOS detected by Jiang et al. [15]. At the same time, the levels of FBG, FINS, HOMA-IR, sex hormones (E2 level was lower than that in the control group) and TC in PCOS group were significantly higher than those in the control group, suggesting that the abnormality of miR-363-3p might be involved in the pathogenesis of PCOS and related to the blood biochemical abnormality of PCOS patients. In 1980, Burghen described the relationship among hyperinsulinemia, insulin resistance (IR) and PCOS for the first time. Statistics show that about 50-70% of PCOS patients would develop into IR [22]. The FBG level of PCOS patients is generally higher than that of the control group. Longterm high-level of FBG can make the blood insulin content increase continuously, thus inducing hyperinsulinemia, further inducing insulin resistance, and ultimately inducing the disorder of glucose metabolism and reproductive hormones [23]. Presently, the correlation between miR-363-3p and insulin resistance has only been verified in animal experiments. Shu et al. found that miR-363-3p expression was significantly reduced in a mouse model of high-fat diet-induced insulin resistance, while resveratrol injection significantly enhanced the expression level of miR-363-3p in the mouse [24]. On this basis, the correlation between miR-363-3p and clinical indicators in PCOS patients was further analyzed, and then it was found that the levels of FBG, FINS, HOMA-IR, sex hormones, AMH and TC were negatively correlated with the level of miR-363-3p, which once again proved the important role of miR-363-3p in PCOS.

Due to rare ovulation, PCOS patients have a low probability of natural pregnancy, so ovulation induction therapy is needed to meet fertility requirements [25]. In this study, after receiving ovulation induction therapy, 70 PCOS patients were successfully pregnant, and 56 others were not pregnant. After further analysis, it was found that compared with the control group, the serum levels of miR-363-3p in pregnant and non-pregnant patients showed a downward trend, while the miR-363-3p levels of non-pregnant group decreased more than that in the pregnant group. Logistic regression analysis suggested that high levels of LH, T, PRL and low level of miR-363-3p in PCOS patients may be independent risk factors for pregnancy failure after ovulation induction therapy. These indicators can predict the pregnancy situation of PCOS patients after ovulation induction therapy to some extent. Furthermore, after a year's follow-up, it was observed that the proportion of premature delivery, fetal macrosomia, gestational diabetes was significantly higher in PCOS patients than in healthy women. This result may be due to abnormal hormone levels in PCOS patients, which lead to an abnormal uterine environment. Although some PCOS patients can get pregnant, embryo development may also be adversely affected, which requires patients and medical workers to pay more attention and closely monitor the specific conditions of pregnant women and fetuses [26].

There are several limitations in this study that need to be explored. First, studies have found that inflammatory reaction is significantly correlated with the occurrence and development of PCOS [27, 28]. This study did not analyze the inflammatory indicators of the subjects, so the relationship between miR-363-3p and inflammation cannot be verified for the time being. This issue needs to be explored by subsequent experiments. Secondly, this study did not explore the mechanism of miR-363-3p regulating PCOS, for example, the effect of miR-363-3p on ovarian granulosa cells. Thirdly, this study did not analyze the relationship between miR-363-3p and endometrial receptivity. Since endometrial receptivity is related to embryo implantation, this part of the study is indeed important. Finally, in this study, the expression of miR-363-3p in the subjects' follicular fluid was not collected for analysis, so the relationship between miR-363-3p in peripheral blood and in follicular fluid could not be evaluated at present. In view of the above problems, we believe that in the future scientific research work, we should collect more related literature reports, by referring to the previous research ideas, we can find our own shortcomings and improve the rigor and science of experimental design.

Conclusions

In this paper, it is considered that the decreased expression of serum miR-363-3p and hormone abnormalities are the characteristics of serological examination in PCOS patients. MiR-363-3p and LH, T, PRL may be the risk factors affecting pregnancy failure in PCOS patients after ovulation induction therapy.

Acknowledgements

Not applicable.

Author Contribution

MS conceived and designed the experiments. YY, ZZ and YQZ performed the experiments, contributed sample collection and statistical analysis. MS wrote the manuscript. HPC contributed with interpretation of the results and critically revised the manuscript. The authors read and approved the final manuscript.

Funding

This work was supported by 2022 Huanggang Municipal Science and Technology Innovation General Project (Grant numbers: YBXM20220005).

Data Availability

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

This study was performed in line with the principles of the Declaration of Helsinki. The experimental scheme complies with the requirements of the Ethics Committee of Huanggang Central Hospital of Yangtze University. The study is approved by the Ethics Committee of Huanggang Central Hospital of Yangtze University, and the patients and their families signed the informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 30 December 2022 / Accepted: 8 April 2023 Published online: 15 May 2023

References

 Wang F, Wang S, Zhang Z, Lin Q, Liu Y, Xiao Y, et al. Defective insulin signaling and the protective effects of dimethyldiguanide during follicular development in the ovaries of polycystic ovary syndrome. Mol Med Rep. 2017;16(6):8164–70.

- Ma Y, Jin J, Tong X, Yang W, Ren P, Dai Y, et al. ADAMTS1 and HSPG2 mRNA levels in cumulus cells are related to human oocyte quality and controlled ovarian hyperstimulation outcomes. J Assist Reprod Genet. 2020;37(3):657–67.
- Chen Z, Zhang M, Qiao Y, Yang J. Effects of letrozole in combination with low-dose intramuscular injection of human menopausal gonadotropin on ovulation and pregnancy of 156 patients with polycystic ovary syndrome. Pak J Med Sci. 2016;32(6):1434–8.
- Liu M, Zhu H, Hu X, Zhu Y, Chen H. Efficacy of different forms of Guizhi Fuling Wan on reproduction and metabolism in women with polycystic ovary syndrome: a protocol for a meta-analysis of randomized controlled trials. Med (Baltim). 2020;99(44):e22954.
- Saei Ghare Naz M, Ramezani Tehrani F, Behroozi-Lak T, Mohammadzadeh F, Kholosi Badr F, Ozgoli G. Polycystic ovary syndrome and pelvic floor dysfunction: a narrative review. Res Rep Urol. 2020;12:179–85.
- Guan S, Zhang Z, Wu J. Non-coding RNA delivery for bone tissue engineering: Progress, challenges, and potential solutions. iScience. 2022;25(8):104807.
- Bongrani A, Mellouk N, Rame C, Cornuau M, Guerif F, Froment P et al. Ovarian Expression of Adipokines in Polycystic Ovary Syndrome: A Role for Chemerin, Omentin, and Apelin in Follicular Growth Arrest and Ovulatory Dysfunction?Int J Mol Sci. 2019;20(15).
- Li W, Zhang T, Guo L, Huang L. Regulation of PTEN expression by noncoding RNAs. J Exp Clin Cancer Res. 2018;37(1):223.
- 9. Gebremedhn S, Ali A, Hossain M, Hoelker M, Salilew-Wondim D, Anthony RV et al. MicroRNA-Mediated Gene Regulatory Mechanisms in Mammalian Female Reproductive Health.Int J Mol Sci. 2021;22(2).
- Zhou H, He Q, Li C, Alsharafi BLM, Deng L, Long Z, et al. Focus on the tumor microenvironment: a seedbed for neuroendocrine prostate cancer. Front Cell Dev Biol. 2022;10:955669.
- Hsu KW, Wang AM, Ping YH, Huang KH, Huang TT, Lee HC, et al. Downregulation of tumor suppressor MBP-1 by microRNA-363 in gastric carcinogenesis. Carcinogenesis. 2014;35(1):208–17.
- Sun Q, Zhang J, Cao W, Wang X, Xu Q, Yan M, et al. Dysregulated miR-363 affects head and neck cancer invasion and metastasis by targeting podoplanin. Int J Biochem Cell Biol. 2013;45(3):513–20.
- Mir FA, Mall R, Iskandarani A, Ullah E, Samra TA, Cyprian F, et al. Characteristic MicroRNAs linked to dysregulated metabolic pathways in qatari adult subjects with obesity and metabolic syndrome. Front Endocrinol (Lausanne). 2022;13:937089.
- Liu Y, Fan L, Jin L, Lu C, Li T, Zhang Z, et al. Integrated bioinformatic analysis of dysregulated microRNA-mRNA co-expression network in ovarian endometriosis. Acta Obstet Gynecol Scand. 2022;101(10):1074–84.
- Jiang X, Li J, Zhang B, Hu J, Ma J, Cui L, et al. Differential expression profile of plasma exosomal microRNAs in women with polycystic ovary syndrome. Fertil Steril. 2021;115(3):782–92.
- Yalcin Bahat P, Ozel A, Demirci A. Evaluation of carotid artery intima-media thickness as a Cardiovascular risk factor in patients with polycystic ovary syndrome. Cureus. 2021;13(1):e13025.
- 17. Zhou F, Li C, Zhang SY. NLRP3 inflammasome: a new therapeutic target for high-risk reproductive disorders? Chin Med J (Engl). 2020;134(1):20–7.
- Javed Z, Papageorgiou M, Deshmukh H, Kilpatrick ES, Mann V, Corless L et al. A Randomized, Controlled Trial of Vitamin D Supplementation on Cardiovascular Risk Factors, Hormones, and Liver Markers in Women with Polycystic Ovary Syndrome. Nutrients. 2019;11(1).
- Yurtdas G, Akdevelioglu Y. A New Approach to Polycystic Ovary Syndrome: the gut microbiota. J Am Coll Nutr. 2020;39(4):371–82.
- 20. Hammond SM. An overview of microRNAs. Adv Drug Deliv Rev. 2015;87:3-14.
- 21. Chen Y, Gao DY, Huang L. In vivo delivery of miRNAs for cancer therapy: challenges and strategies. Adv Drug Deliv Rev. 2015;81:128–41.
- 22. Jabbour R, Ott J, Eppel W, Frigo P. Carotid intima-media thickness in polycystic ovary syndrome and its association with hormone and lipid profiles. PLoS ONE. 2020;15(4):e0232299.
- Chen Y, Zhang W, Chen J, Wang N, Chen C, Wang Y, et al. Association of Phthalate exposure with thyroid function and thyroid homeostasis parameters in type 2 diabetes. J Diabetes Res. 2021;2021:4027380.
- 24. Shu L, Zhao H, Huang W, Hou G, Song G, Ma H. Resveratrol Upregulates mmu-mir-363-3p via the PI3K-Akt pathway to improve insulin Resistance Induced by a High-Fat Diet in mice. Diabetes Metab Syndr Obes. 2020;13:391–403.
- 25. Tang ZR, Zhang R, Lian ZX, Deng SL, Yu K. Estrogen-Receptor Expression and Function in Female Reproductive Disease.Cells. 2019;8(10).

- 26. Cardinale V, Lepore E, Basciani S, Artale S, Nordio M, Bizzarri M et al. Positive Effects of alpha-Lactalbumin in the Management of Symptoms of Polycystic Ovary Syndrome.Nutrients. 2022;14(15).
- Zheng YH, Xu Y, Ma HX, Liang CJ, Yang T. Effect of High-Fat Diet on the Intestinal Flora in Letrozole-Induced polycystic ovary syndrome rats. Evid Based Complement Alternat Med. 2021;2021:6674965.
- Szczuko M, Zapalowska-Chwyc M, Maciejewska D, Drozd A, Starczewski A, Stachowska E. Significant improvement selected mediators of inflammation

in phenotypes of women with PCOS after reduction and low GI Diet. Mediators Inflamm. 2017;2017:5489523.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.